

6 β -HYDROXYIPOLAMIIDE, AN IRIDOID GLUCOSIDE FROM *STACHYTARPHETA MUTABILIS*

CARLO DE LUCA, MARCELLA GUISO*† and CARMELA MARTINO†

Centro di Studio CNR per l'Elettrochimica e la Chimica Fisica delle Interfasi, Via del Castro Laurenziano n. 7, 00161 Roma, Italy,

†Centro di Studio CNR per la Chimica delle Sostanze Organiche Naturali, Istituto di Chimica Organica dell'Università, P.le Aldo Moro n. 2, 00185 Roma, Italy

(Revised received 13 September 1982)

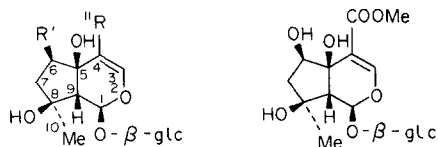
Key Word Index—*Stachytarpheta mutabilis*, Verbenaceae, iridoid glucosides, 6 β -hydroxyipolamiide

Abstract—A new iridoid glucoside has been isolated from *Stachytarpheta mutabilis* and assigned the structure and configuration of 6 β -hydroxyipolamiide on the basis of ^1H NMR and ^{13}C NMR evidence. The conversion of this compound into penta-acetylamiol proved the above assignment.

INTRODUCTION

Stachytarpheta mutabilis Jacq. Vahl (Verbenaceae), a shrub growing in tropical America and Africa, is used in the traditional medicine of East Africa to cure headache, dysentery and eye infections [1]. In the Caribbean area *Stachytarpheta* species are considered as medicinal plants in traditional medicine due to their antifever and anti-dysenteric properties.

In the *Stachytarpheta* genus the presence of iridoid glucosides, mainly ipolamiide (1) [2], has been established [3, 4]. However 1 was isolated only recently from *S. mutabilis* [5]. A chromatographic examination of the ethanolic extract of this plant revealed the presence of ipolamiide (1), the most abundant iridoid component, and of one other compound with a probable iridoid structure. We now demonstrate the structure of this compound to be 6 β -hydroxyipolamiide (2).



1 R = COOMe, R' = H

4 R = Me, R' = OH

2

RESULTS AND DISCUSSION

Compound 2 was crystalline with mp 192–193°, $[\alpha]_D = -161^\circ$ and molecular formula $\text{C}_{17}\text{H}_{26}\text{O}_{12}$. Its UV spectrum showed an intense absorption band at 231 nm ($\log \epsilon$, 4.0) and the IR spectrum revealed bands at 1710 and 1645 cm^{-1} . These data show the presence of a conjugated carbonyl function. Acid hydrolysis of 2 gave glucose and tars arising from extensive decomposition of the aglycone. Acetylation of 2 under mild conditions afforded the penta-acetate (3) which still showed hydroxy bands in the IR spectrum. Therefore, in the aglycone moiety of 2 there must be one hydroxyl group which is readily acetylated and one which is difficult to acetylate.

The ^1H NMR spectrum of 2 (see Experimental) revealed an iridoid structure and showed the presence of a carbomethoxy group at C-4, as supported by the chemical shift value of H-3, and of a methyl geminal with a hydroxyl function at C-8. The signals of H-3 and H-9, both singlets, excluded the presence of a proton at C-5. The presence of an ABX system suggested that a methylene group adjacent to a secondary alcoholic function must be present in the cyclopentane ring. Finally, the doublet at δ 4.80, $J = 7.0$ Hz, was due to the glucose β -anomeric proton.

On the basis of the above data, 2 appeared to have the structure of hydroxyipolamiide, but it was different from the lamiide (7 β -hydroxyipolamiide) which was the only known isomer [2]. The decoupled and SFORD ^{13}C NMR spectra of 2 (Table 1) confirmed the suspected structure and the comparison with model iridoids allowed

*To whom correspondence should be addressed

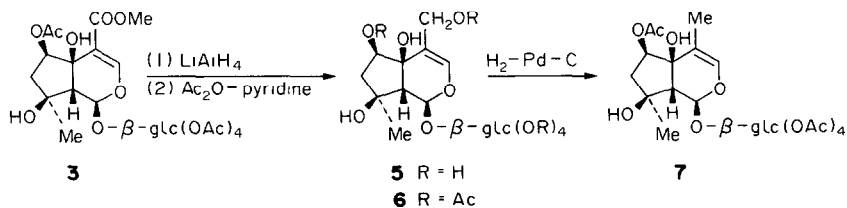


Table 1 ^{13}C NMR spectral data of compounds **1**, **2** and **4** (20 MHz, D_2O , dioxane as internal standard)

Carbon No	1 *	2	4 *
1	94.5 d	94.2 d	93.1 d
3	152.8 d	154.5 d	136.1 d
4	114.0 s	112.8 s	114.6 s
5	71.4 s	70.4†s	72.6 s
6	38.0 t	74.5 d	73.9 d
7	39.4 t	47.0 t	46.8 t
8	78.9 s	74.7 s	75.8 s
9	60.7 d	59.1 d	58.9 d
10	22.8 q	23.6 q	23.8 q
11	169.1 s	169.0 s	11.9 q
OMe	52.5 q	52.7 q	—
1'	99.2 d	99.3 d	98.7 d
2'	73.3 d	73.3 d	73.3 d
3'	76.2 d‡	76.1 d‡	76.2 d‡
4'	70.5 d	70.4 d	70.5 d
5'	77.2 d‡	77.2 d‡	77.0 d‡
6'	61.6 t	61.5 t	61.5 t

*See ref. [6].

†The C-5 signal appears as a singlet in SFORD in the middle of the C-4' doublet

‡These values are interchangeable within each column

the identification of **2** as 6 β -hydroxyipolamide. In fact the chemical shift value of the C-10 methyl (δ 23.6) was very similar to those of compounds like harpagide (δ 24.7) [6], lamiol (**4**) (δ 23.8) and shanzishide methyl ester (δ 24.2) [6] all having a β -hydroxyl function at C-6, a methylene group at C-7 and a β -hydroxyl and an α -methyl at C-8. Comparison of the ^{13}C NMR spectra of **2**, ipolamide (**1**) and lamiol (**4**) (Table 1) shows that the signal of C-8 is more deshielded in **1** than in either **2** or **4** owing to the lack of the γ -effect of the C-6 hydroxyl and that of C-5 appears slightly more deshielded in **1** than in **2** as found for other iridoid glucosides having at C-5/C-6 a β -*cis*-diol function. With lamiol (**4**) C-5 resonates at δ 72.6 while in 5-deoxylamiol it is at δ 74.8 [7] and in 6-deoxylamiol at δ 73.8 [8]. This behaviour is in accord with a *cis*-diaxial interaction between the two hydroxyl functions [9].

The structure of 6 β -hydroxyipolamide proposed for **2** was also in accordance with the chemical shift value, in the ^1H NMR spectrum of **2**, of the X part of the ABX system, in that this signal appeared deshielded owing to the presence of the C-11 carbomethoxy group. A comparison of the ^1H NMR spectra of **2** and **4** [10] showed for the H-6 of **2** a deshielding of δ 0.25; this value is very close to that found in the related compounds, shanzishide methyl ester [11] and 5-deoxylamiol [7], which are the 5-deoxy derivatives of **2** and **4**, respectively.

To obtain chemical evidence that **2** had the structure and configuration of 6 β -hydroxyipolamide, we treated **3** with lithium aluminium hydride to prepare 6 β -hydroxyipolamidol (**5**). This latter compound was transformed by acetylation under mild conditions into its hexa-acetate (**6**) which, by allylic hydrogenolysis with $\text{H}_2/\text{Pd}-\text{C}$, afforded a penta-acetate found to be identical to penta-acetylamiol (**7**) [10].

EXPERIMENTAL

CC: Si gel 70–230 mesh (Merck), cellulose CF₁₁ (Whatman), TLC: Si gel 60 F₂₅₄ and cellulose pre-coated plates (Merck), PC: Schleicher and Schull No. 2043 b Mgl paper, Spray reagents: 2 N H_2SO_4 , vanillin (2 g vanillin, 4 ml conc. HCl, 100 ml MeOH), removal of all volatile materials was performed under red pres.

Isolation of iridoids *S. mutabilis* was collected in Guataparo near Valencia (Estado Carabobo, Venezuela). Voucher specimens of this plant were identified by Dr. Victor M. Badillo, at the Universidad Central de Venezuela, Maracay, and deposited in the herbarium (labelled Carlo De Luca No. 10).

Dried plant material (1 kg) was roughly chopped and extracted with EtOH (6 \times 0.5 l) at room temp. by percolation. PC with *n*-BuOH–HOAc– H_2O (63:10:27) showed at least four spots: R_f 0.51 unknown X, 0.37 **1**, 0.25 **2** and 0.18 unknown Y. The ethanolic extracts were concd to a viscous fluid, then distributed between H_2O and EtOAc and the aq. layer further extracted (twice) with EtOAc. The resulting aq. soln was concd and treated with decolorizing charcoal (300 g). The suspension was stratified on a Gooch funnel (14 cm i.d.). Monosaccharides were eluted with H_2O (10 l), di- and oligosaccharides with 5% EtOH (1 l), 10% EtOH (1 l), **1**, **2** and unknown Y with 30% EtOH (5 l) and 50% EtOH (2 l) (fraction A), small amounts of **1**, other compounds and unknown X with 70% EtOH (2 l) (fraction B). Fraction A (8 g) was chromatographed on Si gel in *n*-BuOH satd with H_2O to give **1** (3 g), and **2** (0.5 g) still contaminated with **1**. This latter was rechromatographed as above to give **2** (0.35 g) which was twice crystallized from MeOH–Me₂CO, mp 192–193°, $[\alpha]_D^{25} - 161^\circ$ (MeOH, c 0.2), UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm (log ϵ): 231 (4.0), IR $\nu_{\text{max}}^{\text{KBr}}$ cm^{-1} : 1710, 1645, 1395, 1310, 1080. ^1H NMR (90 MHz, D_2O) δ 7.60 (1H, s, H-3), 5.86 (1H, s, H-1), 4.15 (1H, *pt*, H-6, X part of ABX system), 3.77 (3H, s, COOMe-11), 2.63 (1H, s, H-9), 2.16 and 1.90 (2H, AB part, $J_{\alpha-\beta} = 13.5$, $J_{\gamma-\delta} = 8.0$, $J_{\beta-\delta} = 6.4$ Hz, 2H-7), 1.18 (3H, s, 3H-10) (Found: C, 48.20, H, 6.32. $\text{C}_{17}\text{H}_{26}\text{O}_{12}$ requires: C, 48.34, H, 6.20%).

Penta-O-acetyl derivative (3) of 2. Compound **2** (50 mg) was treated with dry pyridine (0.4 ml) and Ac₂O (0.8 ml) for 1 hr at room temp. After addition of MeOH (3 ml) the soln was left for 30 min, then evaporated to give crude **3** which, when chromatographed on Si gel in Et₂O–EtOAc (6:4), gave pure **3** (55 mg). Needles from MeOH, mp 202.5–203.5°, IR $\nu_{\text{max}}^{\text{CHCl}_3}$ cm^{-1} : 3650, 3500, 1760, 1645, 1445, 1380, 1310, 1260, 1130, 1075, 1045, 960, 920, 875. ^1H NMR (90 MHz, CDCl_3) δ 7.40 (1H, s, H-3), 5.63 (1H, *d*, $J_{1-2} = 3.0$ Hz, H-1), 5.49 (1H, *pt*, H-6, X part of ABX), 3.75 (3H, s, COOMe-11), 2.71 (1H, *d*, $J_{8-9} = 3.0$ Hz, H-9), 1.29 (3H, s, 3H-10).

Reduction of 3 to yield 7 β -hydroxyipolamidol (5). LiAlH₄ (45 mg) suspended in dry THF (3 ml) was added dropwise to a stirred soln of **3** (60 mg) in dry THF (5 ml). After 1 hr at room temp. the suspension was cooled and MeOH was added dropwise to destroy the excess LiAlH₄. The resulting suspension was diluted with H_2O (10 ml) and CO₂ was bubbled through the soln until pH ca 8. Finally it was concd, further treated with CO₂ (pH ca 7) and centrifuged. The residue was washed (\times 5) with H_2O until it gave a negative reaction with the vanillin reagent. The collected aq. solns were treated with decolorizing charcoal (0.5 g) and the suspension was stratified on a Gooch funnel (2 cm i.d.), washed with H_2O (200 ml) and eluted with MeOH (50 ml). The MeOH soln was evaporated to give a residue (30 mg) which, after chromatography on cellulose developed with *n*-BuOH satd with H_2O gave pure **5** (20 mg) as an amorphous powder. ^1H NMR (60 MHz, D_2O) δ 6.25 (1H, *br* s, H-3), 5.51 (1H, s, H-1), 4.10 (2H-11), 2.42 (1H, s, H-9), 1.6–2.2 (2H-7), 1.10 (3H, s, 3H-10).

Acetylation of 5 to give the hexa-acetate (6). Compound **5** (20 mg) was acetylated and worked-up as described for **3** to give

pure **6** (22 mg) ^1H NMR (90 MHz, CDCl_3) 6.43 (1H, *br s*, H-3), 5.64 (1H, *br s*, H-1), 4.64 (2H, *br s*, 2H-11), 2.65 (1H, *br s*, H-9), 1.22 (3H, *s*, 3H-10)

Hydrogenolysis of 6 to yield penta-acetylamiol (7) Compound **6** (22 mg) dissolved in 95% EtOH (5 ml) was added to 10% Pd-C (10 mg) previously suspended in 95% EtOH (3 ml) and satd with H_2 . After 5 min the catalyst was removed by filtration and the soln gave a residue which, after chromatography on Si gel in Et_2O -EtOAc (8/2) gave a compound (6 mg) shown to be identical to penta-acetylamiol (**7**) (mp and mmp identical, IR and ^1H NMR spectra superimposable)

REFERENCES

- 1 Kokwaro J O (1976) *Medicinal Plants of East Africa* p 224 EAL, Nairobi
- 2 Scarpati, M L and Guiso, M (1969) *Gazz Chim Ital* **99**, 1150.
- 3 Kooiman, P (1970) *Acta Bot Neerl* **19**, 329
- 4 Tantisewie, B. and Sticher, O (1975) *Phytochemistry* **14**, 1462.
- 5 De Luca, C (1980) *Fitoterapia* **51**, 279
- 6 Bianco, A, Caciola, P., Guiso, M, Iavarone, C and Trogolo, C (1981) *Gazz Chim Ital.* **111**, 201
- 7 Agostini, A, Guiso, M, Marini-Bettolo, R and Martinazzo, G (1982) *Gazz Chim Ital* **112**, 9
- 8 Guiso, M and Martino, C, *J Nat Prod* (in press).
- 9 Damtoft, S., Jensen, S R and Nielsen, B J (1981) *Phytochemistry* **20**, 2717.
- 10 Scarpati, M. L and Guiso, M (1967) *Tetrahedron* **23**, 4709
- 11 Bianco, A, Francesconi, A and Passacantilli, P (1981) *Phytochemistry* **20**, 1421